



Nanofibrillated cellulose: results of *in vitro* and *in vivo* toxicological assays

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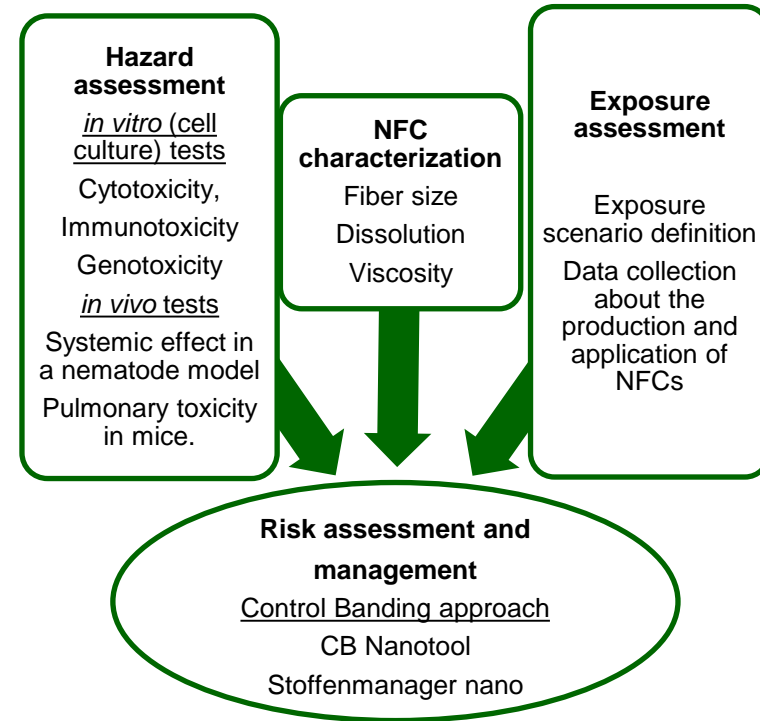
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Hazard assessment approach for NFC

Methodology

- ***In vitro* assays for cytotoxicity, genotoxicity, and immunotoxicity**
- **A nematode model for systemic effects and neurotoxicity**
- **Pharyngeal aspiration study with mice for pulmonary immunotoxicity and genotoxicity**



NFC samples studied

NFC-VTT

- Masuko grinder (5 passages)
- No pre-treatment, no bioside
- **Bacteria**

NFC-CTP/VTT

- Masuko grinder (5 passages)
- Enzymatic pre-treatment, bioside added to the pulp
- **Bacteria**

NFC-TE/VTT

- Masuko grinder (3 passages)
- TEMPO-mediated oxidation as a pre-treatment, no bioside
- **Bacteria and yeast**

NFC-TE/CTP samples studied

NFC-TE/CTP B1

- Lab scale high-pressure homogenizer
- TEMPO-mediated oxidation as pre-treatment
- Dialysed to remove traces of reactant
- No biocide added
- **Bacteria**

NFC-TE/CTP B2 ± biocide Busan 1009

- Pilot scale high-pressure homogenizer
- TEMPO-mediated oxidation as pre-treatment
- Washed four times but might contained residues of reagents (TEMPO, sodium bromide and sodium hypochlorite)
- Without biocide: **bacteria, yeast and mold**
- With biocide: **some bacteria**

Cytotoxicity

- **Highest Tolerated Dose (HTD) assay**
 - Qualitative morphological changes in human cervix carcinoma HeLa cells under an inverted phase contrast microscope
 - Comparison with untreated cells and evaluation on a scale ranging from 0 to 4
- **Only marginal effects seen at top doses (→ 2 mg/ml) of NFCs, probably as a result of mechanical influence**
- **Total Protein Content (TPC) assay**
 - A quantitative test measuring growth inhibition of HeLa cells, total protein content as the indicator of the cells' viability
 - Based on reaction of fluorescamine with compounds containing primary amino groups to produce fluorescent products.
- **None of the NFCs indicated a cytotoxic effect**
- **Boar Sperm Motility Assay (BSM)**
 - A qualitative test particularly suitable for testing of suspensions
 - Boar spermatozoa's movement inhibition indicates mitochondrial or membrane damage
- **No toxic effects seen**

Immunotoxicity – pulmonary inflammation

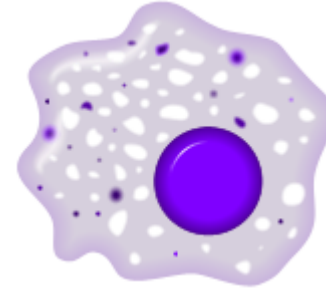
In vitro studies

- Macrophages exposed to several doses of nanocelluloses in cell culture
- **Cytotoxicity** assessed by photometric and luminometric methods
- Macrophage activation assessed by measuring the **expression of essential cytokines**
 - at the mRNA level by real-time quantitative PCR
 - at the protein level by ELISA

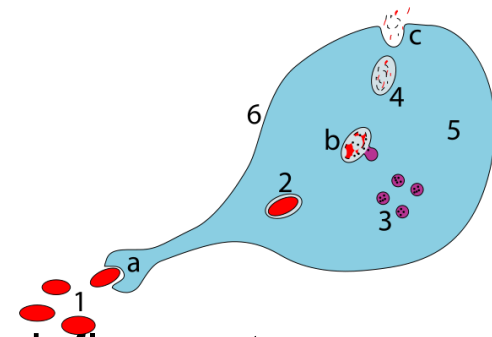
In vivo studies on NFC-TE/CTP B2 + biocide

- Material for *in vivo* studies chosen by the Consortium
- Mice exposed by pharyngeal aspiration
- Inflammatory parameters analysed

Macrophages



- act as the first line of defense against pathogens and particles in the lungs
- ingest pathogens and particles and break them down
- release proinflammatory cytokines which promote inflammatory processes
 - TNF-alpha, IL-1beta, and IL-6 determined in the present study



In the immunotoxicological part of the present study, human macrophages derived from blood monocytes were used

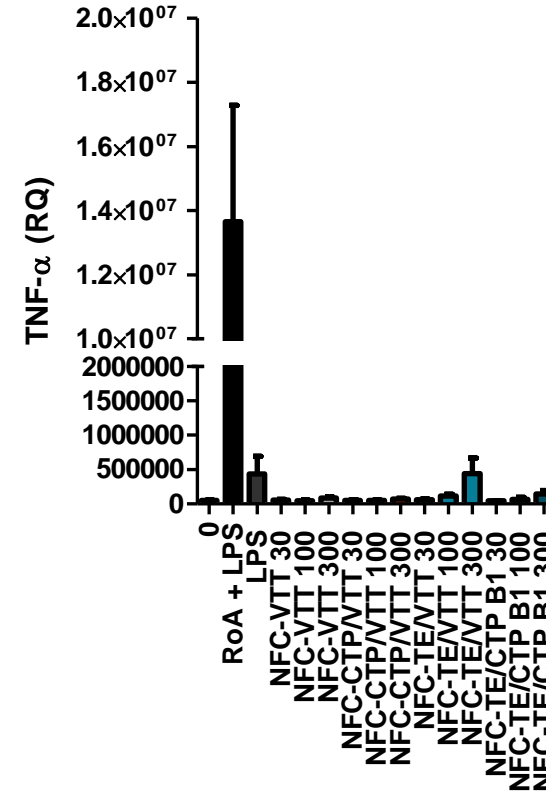
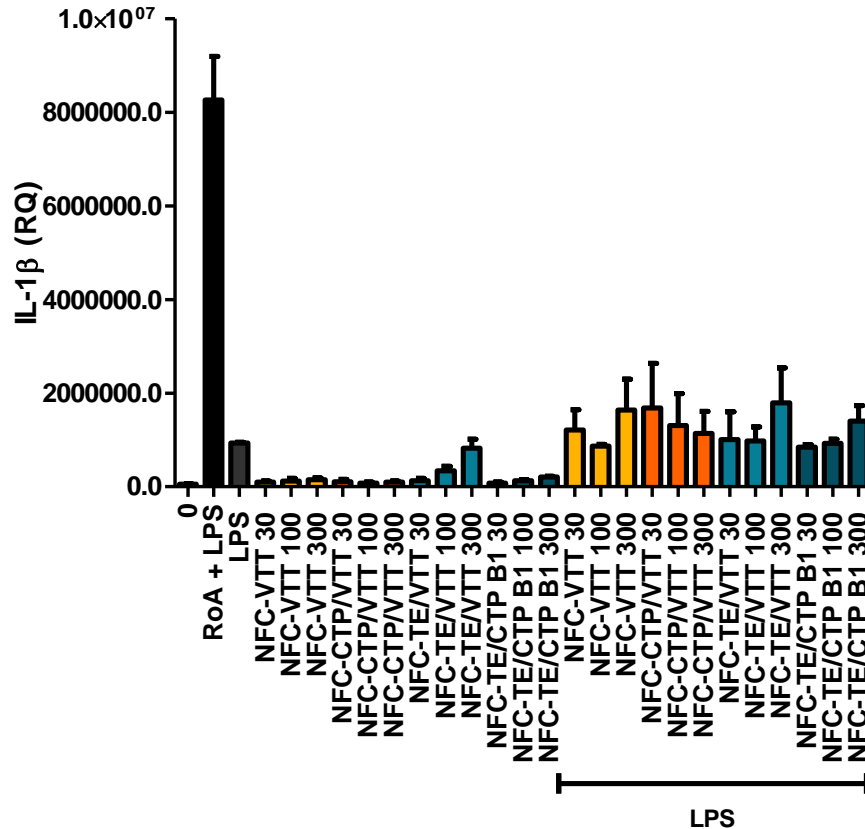
Cytotoxicity in human macrophages

- LDH release from cells expressed as percentage of total LDH, measured photometrically
- ATP in cells expressed as relative luminescence (percent of unexposed control), measured luminometrically
- Human monocyte-derived macrophages exposed for 6 h
- Cell culture supernatants collected
- Cytotoxicity measured by lactate dehydrogenase (LDH) leakage and decrease of ATP (luminescent cell viability assay)
- Roridin A (RoA) and silica used as positive controls

NFCs were not cytotoxic to macrophages at doses up to 300 $\mu\text{g/ml}$

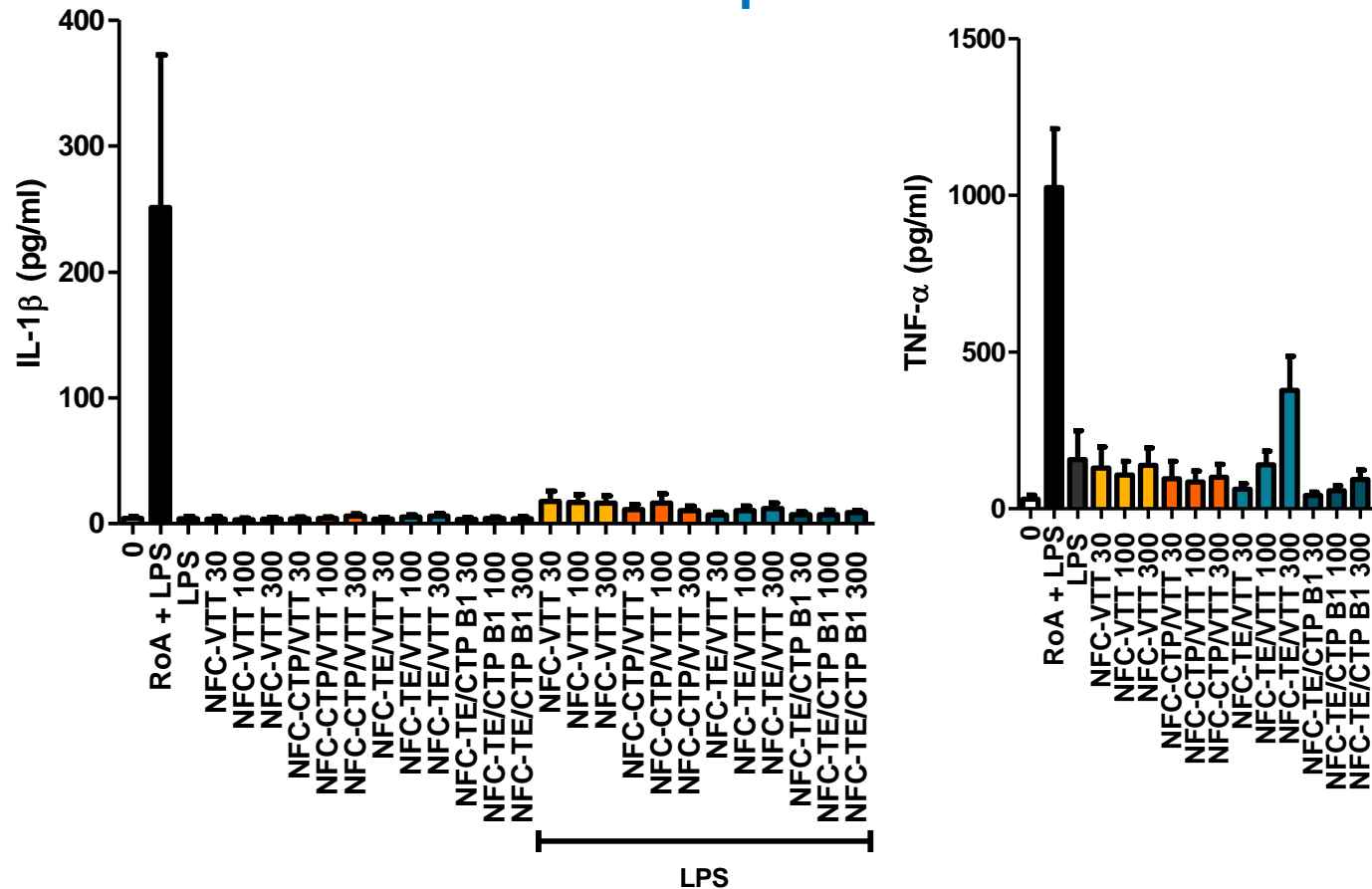
mRNA expression of pro-inflammatory cytokines TNF- α in exposure to NFCs

IL-1 β and



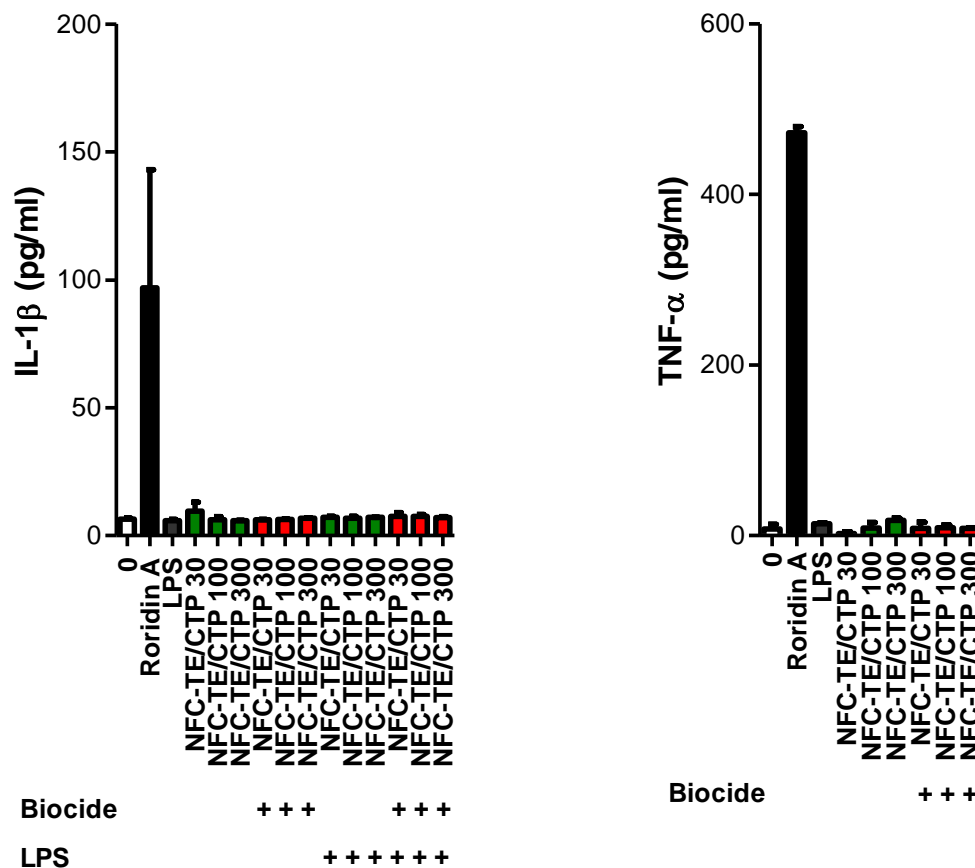
- Primed and unprimed human monocyte-derived macrophages exposed for 6 h
- Cell culture supernatants collected, mRNA measured by PCR
- **NFC-TE/VTT slightly increased IL-1- β and TNF- α mRNA (due to bacteria & yeast?)**

Secretion of pro-inflammatory cytokines IL-1 β and TNF- α in exposure to NFCs



- Primed and unprimed human monocyte-derived macrophages exposed for 6 h
- Cell culture supernatants collected, pro-inflammatory cytokines measured by ELISA
- **All NFCs slightly induced TNF- α and, in LPS-primed samples, IL-1- β**

Secretion of pro-inflammatory cytokines IL-1 β and TNF- α in exposure to NFC-TE/CTP B2 \pm biocide

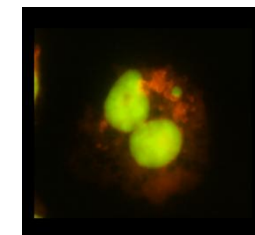
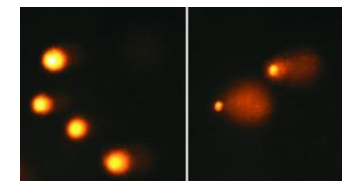
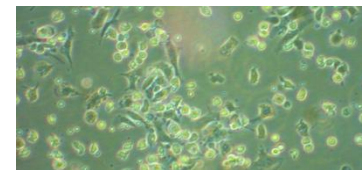


- Primed and unprimed human monocyte-derived macrophages exposed for 6 h
- Cell culture supernatants collected, pro-inflammatory cytokines measured by ELISA
- **No effects observed**

Genotoxicity – identifying possible carcinogens

In vitro studies

- Human bronchial epithelial BEAS 2B cells exposed to several doses of nanocelluloses in cell culture
- **Cytotoxicity** (cell count) utilized for dose finding
- **DNA damage** examined by the Comet assay
- **Oxidative DNA damage** studied by the enzyme-modified Comet assay
- **Chromosome damage** assessed by the micronucleus assay
- **Cell cycle delay** studied by the cytokinesis-block proliferation index



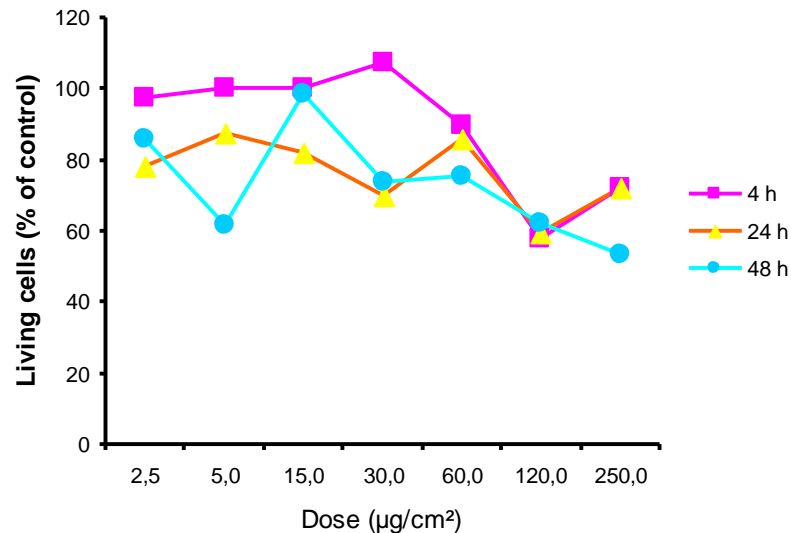
In vivo studies on NFC-TE/CTP B2 + biocide

- One NFC material chosen for *in vivo* studies by the Consortium
- Mice exposed by pharyngeal aspiration
- DNA damage and oxidative DNA damage studied in bronchoalveolar lavage (BAL) cells

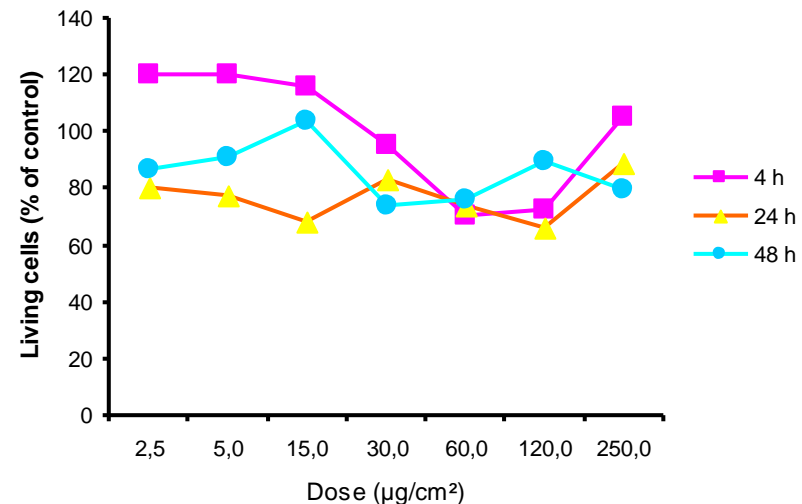
Cytotoxicity of NFC-TE/CTP B2 ± biocide in human bronchial epithelial BEAS 2B cells

- **Double staining:** Propidium iodide stains dead and dying cells, Hoechst stains cell nuclei
- Cell count in fluorescence microscope

NFC-TE/CTP B2 with biocide

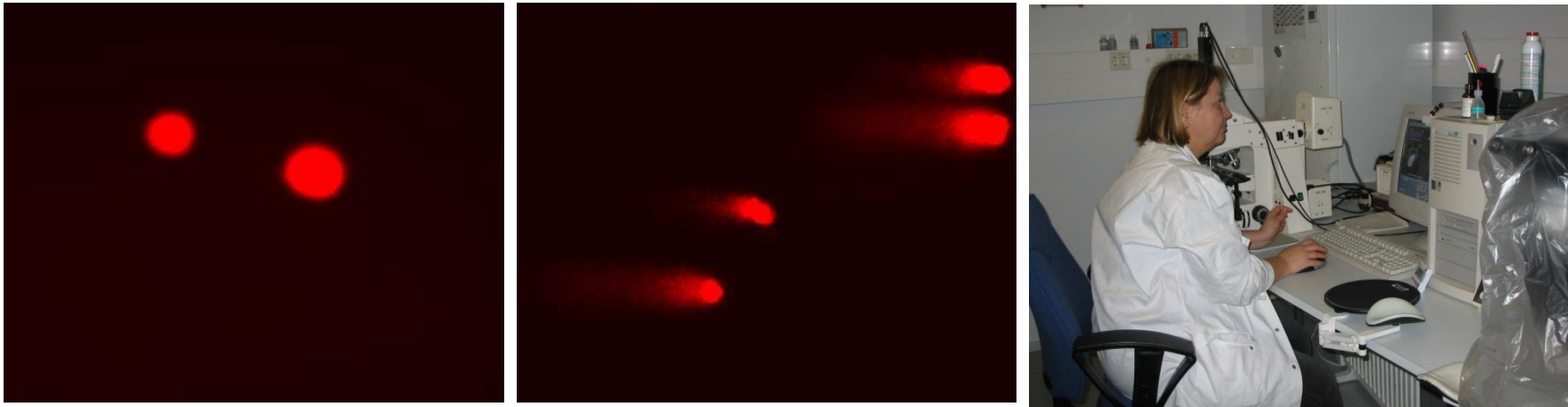


NFC-TE/CTP B2 without biocide



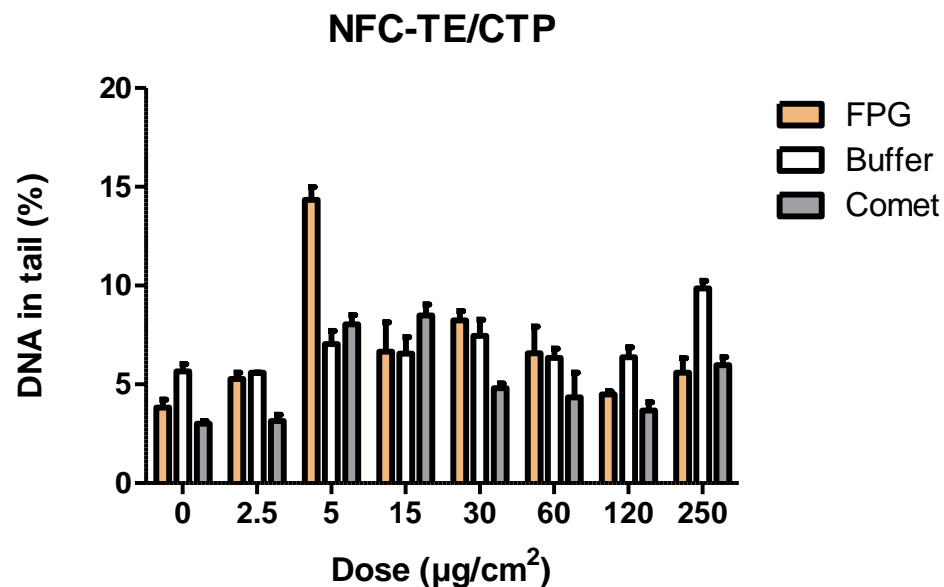
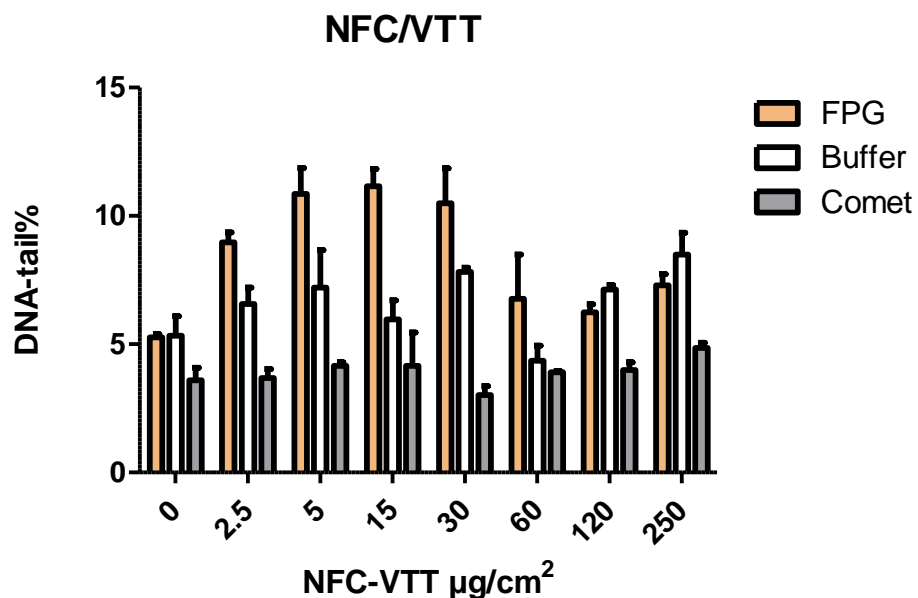
Cytotoxicity of the NFCs in BEAS 2B cells was generally low

DNA damage detection by the comet assay



- DNA with strand breaks wanders out of the nucleus in electrophoresis
- The proportion of DNA in "comet tail" reflects the amount of DNA damage
- Oxidative DNA damage visualized by turning oxidative DNA adducts to strand breaks by a specific enzyme (FPG)
- Analysis in fluorescence microscope using a semiautomatic interactive software

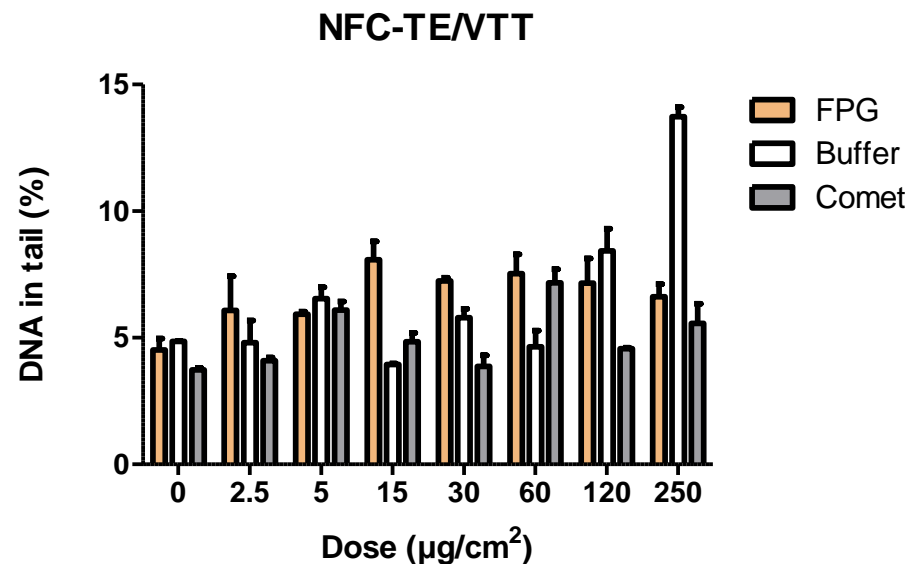
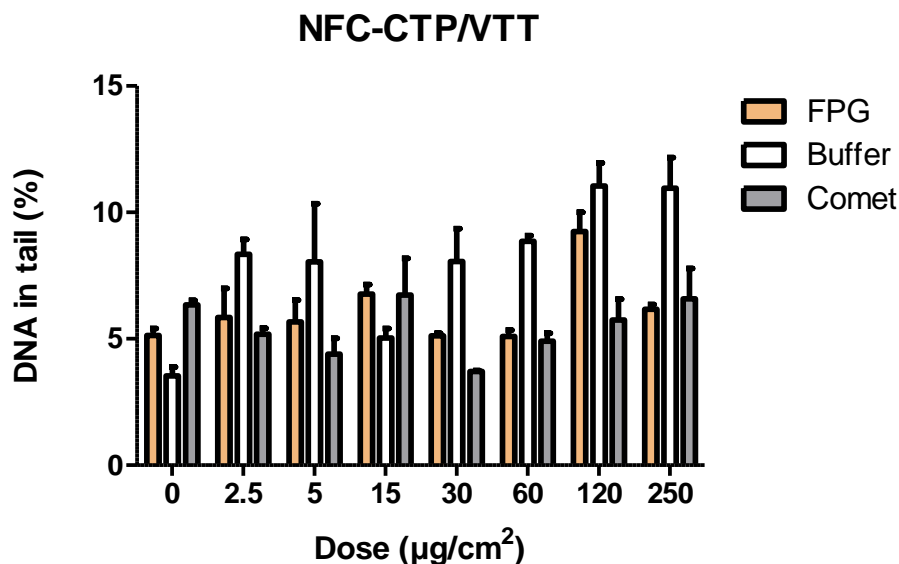
DNA damage induced *in vitro* by NFCs



- **No increase in DNA damage**
- **Slight increase in oxidative DNA damage (FPG vs Buffer)**

- **Slight increase in DNA damage in the comet assay**
- **Slight Increase in oxidative DNA damage at one dose**

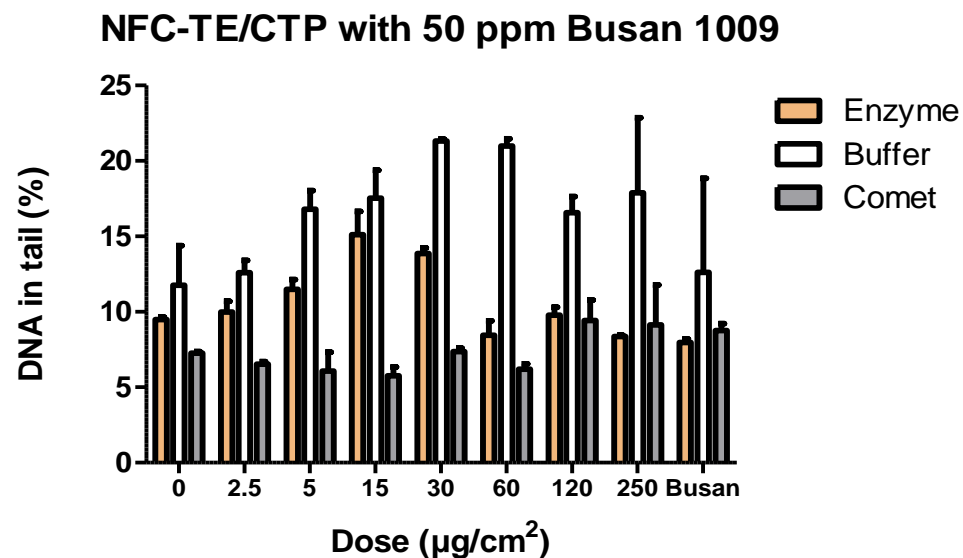
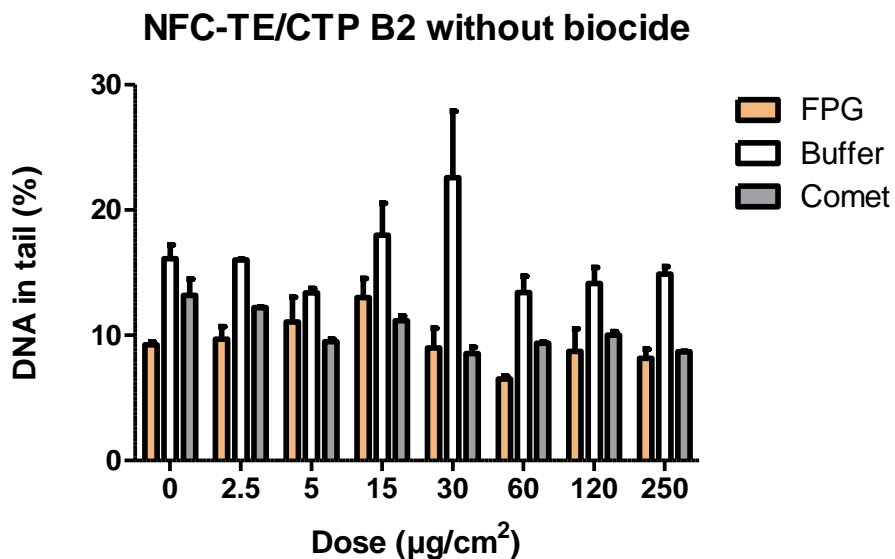
DNA damage induced *in vitro* by NFCs



- Slight Increase in DNA damage in the Buffer and FPG series, **but not in ordinary Comet assay**
- **No increase in oxidative DNA damage**

- Slight increase in DNA damage in the comet assay and Buffer series
- **No increase in oxidative DNA damage**

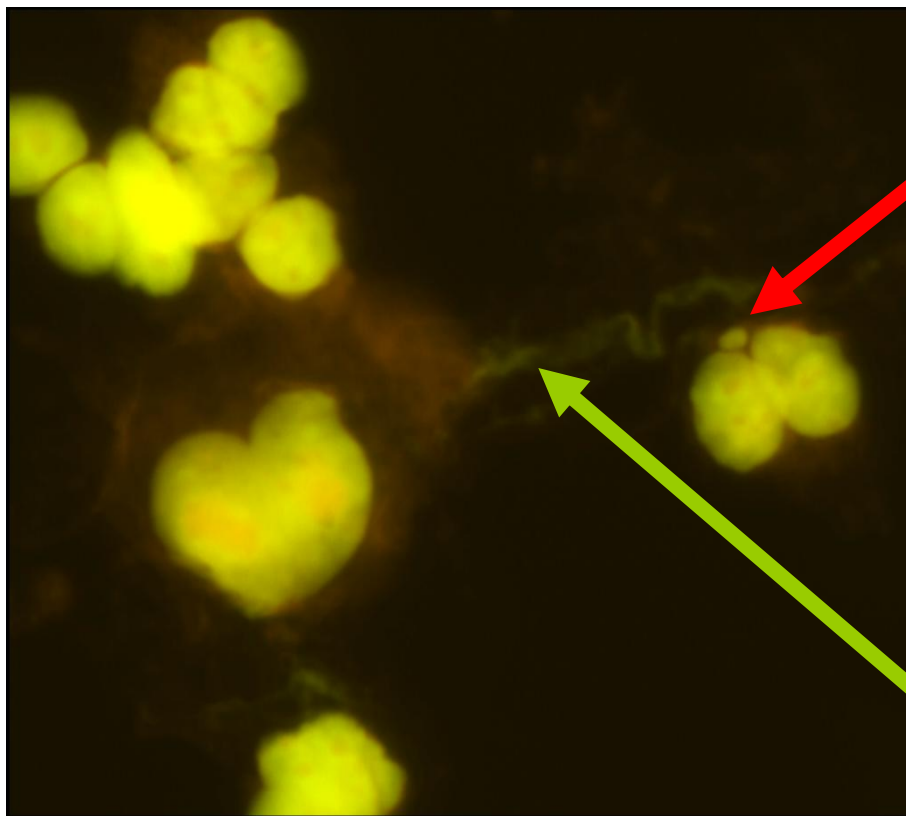
DNA damage by NFC-TE/CTP B2 ± biocide



- No increase in DNA damage
- No increase in oxidative DNA damage

- Increase in DNA damage in the Buffer series, but not in ordinary Comet assay
- No increase in oxidative DNA damage

Micronucleus assay with NFCs



A micronucleus

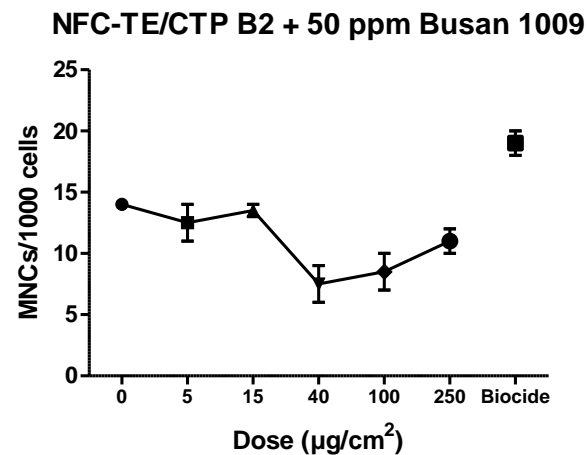
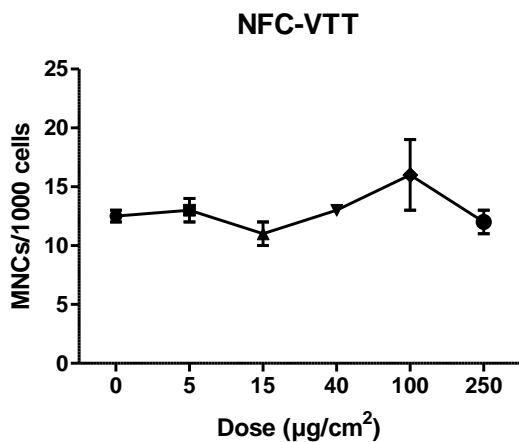
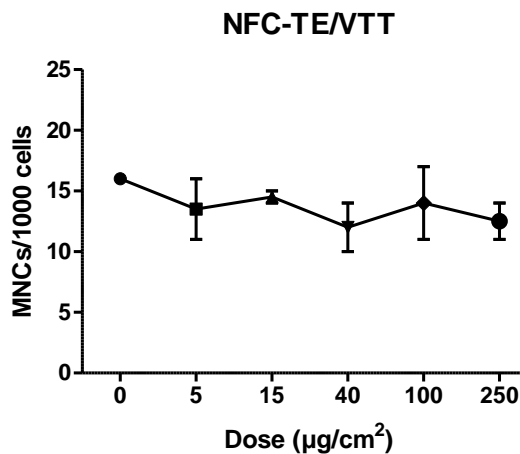
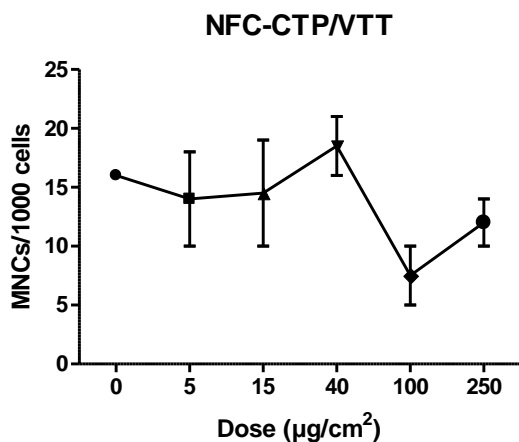
- Micronuclei reflect structural and numerical alterations of chromosomes
- **Acridine orange staining:** nuclei (DNA) are green, cytoplasm (RNA) is red
- Analysis in fluorescence microscope

NFC

Human bronchial epithelial BEAS 2B cells in fluorescence microscope after treatment with NFC-VTT (250 $\mu\text{g}/\text{cm}^2$).

Photo: Kati Hannukainen and Hilikka Järventausta, FIOH

No induction of micronuclei by NFCs in BEAS 2B cells



48-h
treatment

- No effect on cell cycle length either (data not shown)

Nematode toxicity assay *in vivo*

- Transgenic line ($P_{dat-1}::GFP$) of *Caenorhabditis elegans* expressing
- Green Fluorescent Protein in its dopaminergic neurons
- Bright fluorescence of ganglia indicates functional nervous system
- NFC-TE/CTP dose 0.5 mg/ml (biocide)
- Single walled carbon nanotubes used as fibre control, DMSO (5%) as a positive control
- control
- 24-h follow-up for viability, behavior and reproduction
- **The NFCs were not toxic to *C. elegans***

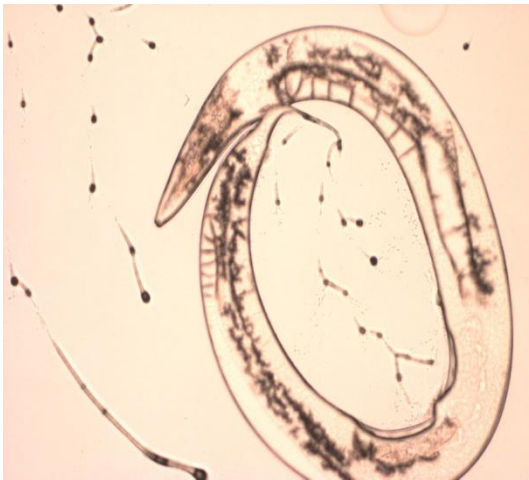


Photo: Prof. Garry Wong,
University of Eastern Finland

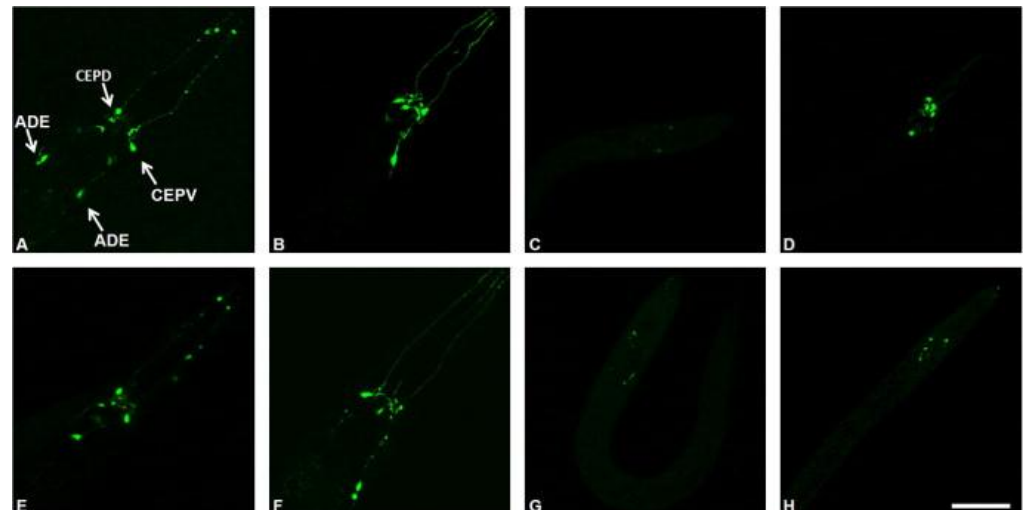
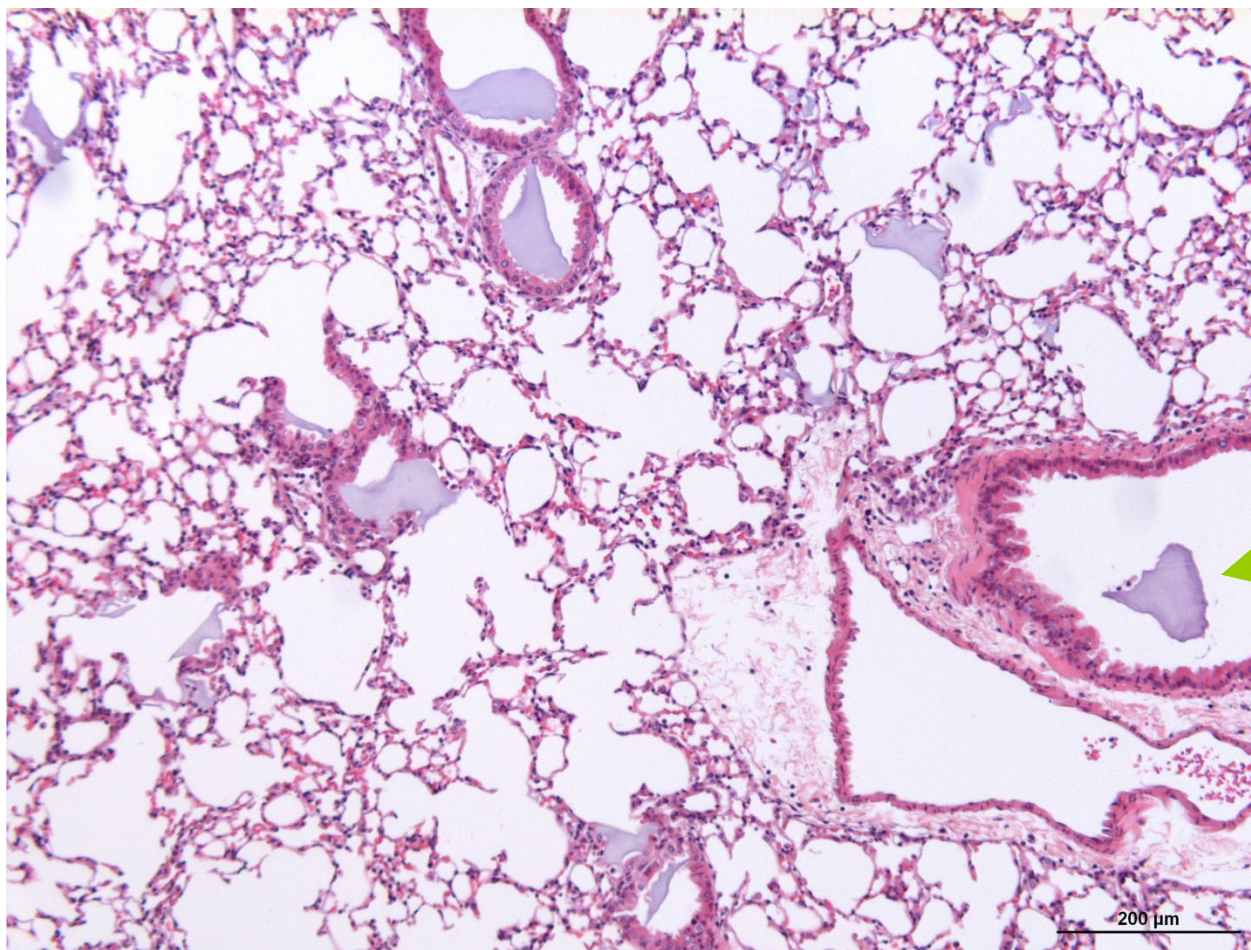


Photo: Jadiya et al. 2011

In vivo study in mice

- Female C57BL/6 mice
- NFC-TE/CTP B1 tested
- Exposure by pharyngeal aspiration
- Single dose: 20, 40, 80 and 200 $\mu\text{g}/\text{mouse}$ (in PBS)
- Biocide Busan 1009 tested separately at the same dose as in the NFC
- Negative and positive controls
- Samples collected 16 h after dosing (acute effect)
 - Bronchoalveolar lavage fluid
 - Lungs
 - Blood
- Analysis of pulmonary inflammation and DNA damage

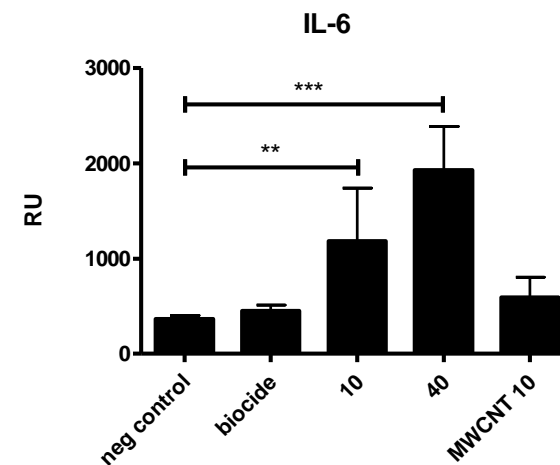
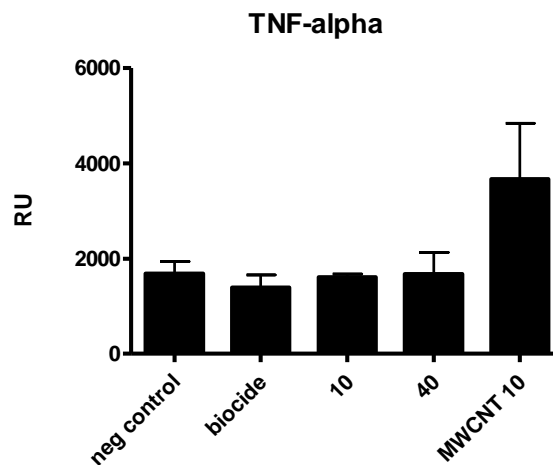
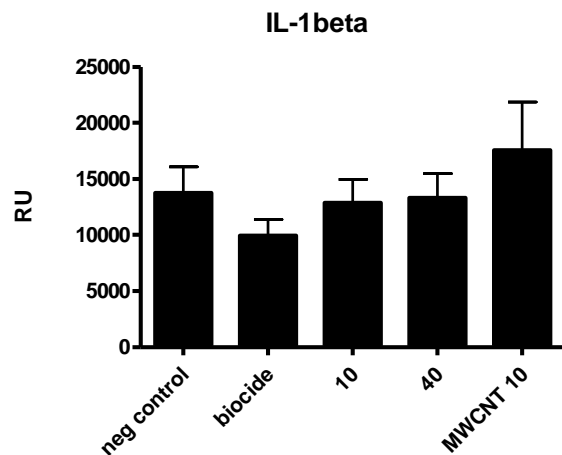
NFC-TE/CTP B2 in mouse lungs



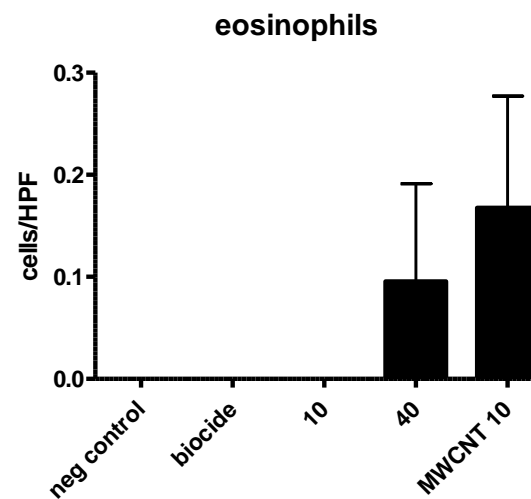
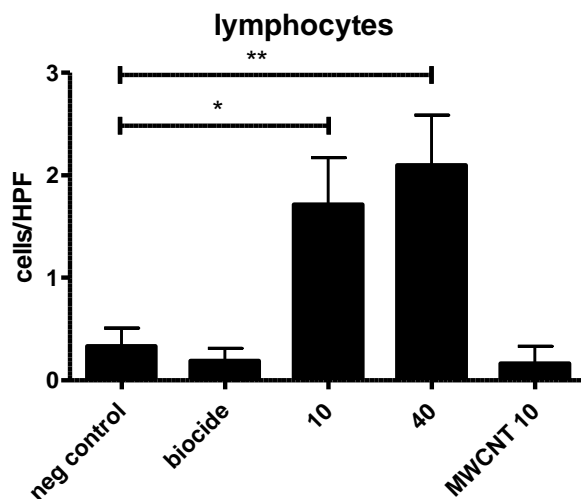
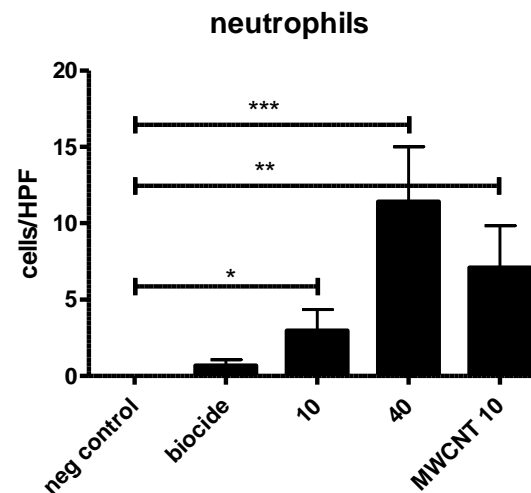
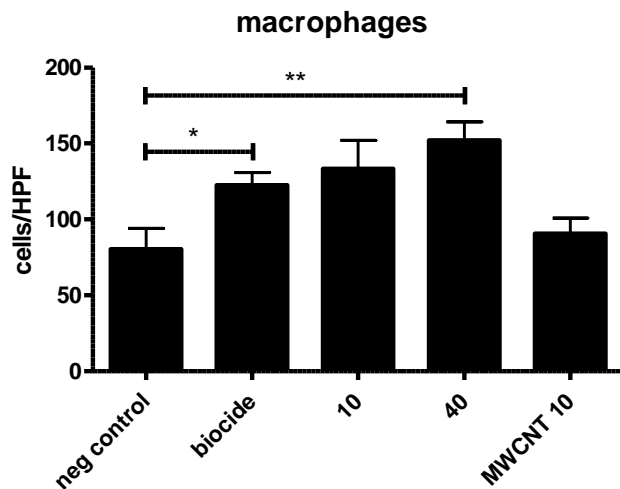
NFC

- In histological samples of the lungs, NFC was seen in or near bronchioles
- No dramatic tissue changes were observed

NFC-TE/CTP B2 induced mRNA of pro-inflammatory cytokine IL-6

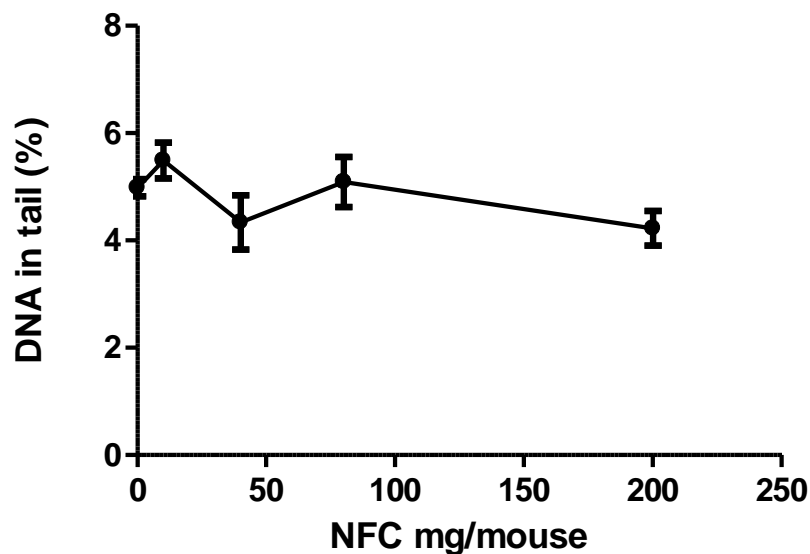


NFC-TE/CTP B2 induced influx of inflammatory cells to mouse lungs

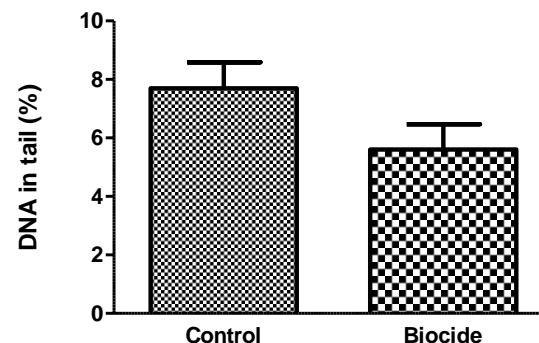


No DNA damage in mouse bronchoalveolar lavage (BAL) cells after pharyngeal aspiration of NFC-TE/CTP B2

DNA damage after pharyngeal aspiration of NFC



DNA damage in biocide-treated mice



Conclusions – toxicity of NFCs

In vitro

- **Low or no cytotoxicity**
- **Slight induction of proinflammatory cytokines** in macrophages with or without LPS priming *in vitro*
- **Slight DNA damage** in human bronchial epithelial cells – similarly to a number of other "inert" nanomaterials
- **Marginal induction of oxidative DNA damage** *in vitro* (some NFCs)
- **No increase in chromosome damage (micronuclei)**

In vivo (NFC-TE/CTP B2, pharyngeal aspiration, mice)

- **No DNA damage** in bronchoalveolar lavage cells
- **Pulmonary inflammation**

→ Possibly due to the particulate / bacteria in the NFC

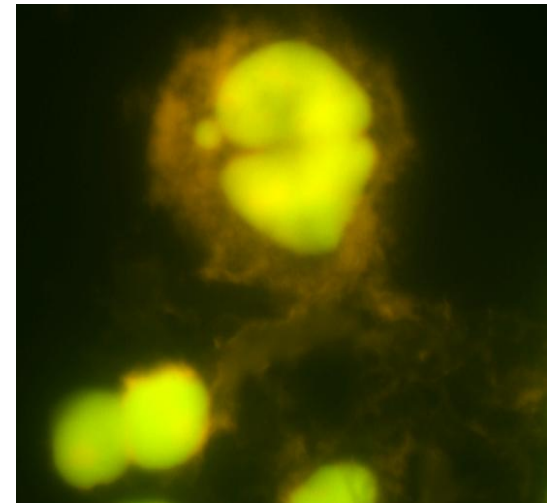
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